

Low intensity pulsed ultrasound in periodontal regeneration

S.Gopalakrishnan¹, Parthiban Saket², V.Shankarram³

¹ Reader,
² Senior lecturer,
³ Professor,
 Thai Moogambigai Dental College and Hospital,
 Mugappair, Chennai

Received : 31.08.2015
 Review Completed : 28.09.2015
 Accepted : 05.10.2015

Introduction

Dental caries and periodontal diseases are the two most diseases of the oral cavity. Not only do they present physical and social hazards but also seriously compromise human health and quality of life. Tissue engineering has contributed to the cessation of many oral diseases. Several methods have been described to enhance cellular performance and low intensity pulsed ultrasound (LIPUS) has shown to play an important role in cell metabolism. LIPUS stimulation is a classical therapeutic modality for bone regeneration and its efficiency has been widely reported over the years. Interestingly, recent studies have provided evidence that LIPUS plays an important role in the metabolism of periodontal cells and tissues as well.

Lipus and Biologic Mechanisms

Normal bone physiology is known to be regulated by mechanical stimulation. Both osteoblastic and osteoclastic activity have been proved to be mediated by mechanical stimuli¹. Osteoclasts initiate bone resorption in response to determined signals and osteoblasts are recruited to deposit bone matrix in response to a coupling signal. Osteoclast and osteoblast activity could thus be related to opposite strain modalities. During orthodontic tooth movement, for example, the networked reactions that occur in and around periodontal ligament and alveolar bone cells change compressive and tensile stresses into molecular events, resulting in bone resorption and formation, respectively². Other types of mechanical stimulation could thus modify cell metabolism, including ultrasound. LIPUS (intensity ranging from 30 - 100 mW/cm²) is an acoustic radiation that can be transmitted into the living tissues as pressure waves resulting in biochemical events at the cellular level³. LIPUS has been shown to stimulate bone and cartilage cells *in vitro*, indicating that ultrasound exerts direct anabolic effects such as production of growth factors and other signaling molecules, osteogenic differentiation and extra cellular matrix production⁴. The mechanisms involved in LIPUS-stimulated tissue repair is by the anabolic biophysical effects caused by mechanical stress and fluid micro-streaming which has an impact on the cellular plasma membrane. This triggers a cascade of intracellular

signal transduction which in turn causes a subsequent gene transcription⁵.

Lipus and Bone Regeneration

Mechanical stimulus to bone is of a great importance for maintaining the bone mass and structural stability of the skeleton. When bone is mechanically loaded, movement of fluid within the spaces surrounding bone cells generates a disparity in fluid levels. This in turn stimulates osteoclasts and osteoblasts, resulting in an enhanced anabolic activity for bone remodeling. Bone repair and regeneration, accelerated bone fracture healing, and an enhanced osteogenesis at the distraction site have been demonstrated during *invivo* studies with the use of Therapeutic LIPUS⁶. LIPUS stimulation has also been shown to enhance expression of bone formation-related genes such as collagen type I and X, aggrecan, transforming growth factor beta, runt related gene-2, osteocalcin⁷, insulin-like growth factor-I, bone sialoprotein and alkaline phosphatase. In addition, LIPUS has been reported to promote protein synthesis and calcium uptake in various osteoblastic cell lines⁸. The enhancement of COX-2 gene expression and synthesis of endogenous prostaglandin E2 (PGE2 plays an important role in bone remodeling⁹.



Lipus and Periodontal Ligament Regeneration

Periodontal ligament is subjected to various kinds of mechanical stress during every stage of cell proliferation and differentiation¹⁰. In a previous study, cyclic stretch stimulation mediated periodontal ligament cells differentiation, thus regulating the function of the periodontal ligament as a source of cementoblasts and osteoblasts through the EGF/EGF-R system¹¹. It has been reported that LIPUS is effective in releasing fibroblast growth factors from a macrophage like cell line¹² and also helps in inducing early cementoblastic differentiation from the periodontal ligament by promoting the formation of substrate and increasing alkaline phosphatase (ALP) activity, and also enables the regeneration of periodontal tissue destroyed by periodontal disease and repair of root resorption¹³. Mostafa et al. (2009) demonstrated that ALP and OPN expressions were also induced in human gingival fibroblasts treated with LIPUS helping in osteogenic potential differentiation¹⁴.

Lipus and Cementum Regeneration

Cementum is a thin mineralized tissue covering the tooth root surface and assists in anchoring teeth to surrounding alveolar bone, maintaining the structural stability and physiological function of the dentition¹⁵. A certain degree of root resorption occurs in most treatment cases, ranging from just a slight apical resorption to a complete tooth root loss¹⁶. The cementum layer covering the root surface plays a

crucial role in preventing resorption during tooth movement. Cementoblasts share many characteristics with osteoblasts, including similar molecular properties and the ability to promote mineralization. Studies have shown that cementum metabolism is also controlled by mechanical stimulus similar to bone. It has been reported that mechanical loading enhances the expression of phenotypic makers such as OCN and BSP in cementoblast however, the expression was just moderately stimulated compared to osteoblasts¹⁷. El-Bialy et al. (2004) have published studies showing that LIPUS prevented root resorption during experimental tooth movement in humans¹⁸. Studies also have showed that LIPUS up-regulated the expression of several genes related to mineral metabolism in mouse cementoblasts¹⁹. LIPUS simulation also significantly up-regulates COX-2 mRNA expression and enhanced PGE2 production inducing cementoblastic differentiation and matrix mineralization through EP2/EP4 prostaglandin receptors pathway²⁰. Furthermore, assessment was done to see the inhibitory effect of LIPUS application on root resorption using an experimental model of tooth replantation involving luxation and immediate replacement of maxillary first molars in rats²¹. The results evidenced that the area of root resorption lacunae was statistically decreased in LIPUS treated sample. In addition, some in vitro studies have shown that LIPUS may contribute to the reduction of the trauma-induced inflammatory reaction through impairment of the TNF-α signaling pathway, suggesting its potential as a therapeutic tool to optimize the regenerative potential of periodontal tissues on replanted teeth²².



Indian Dental Association
Madras Branch

Lipus and Gingival Regeneration

LIPUS application in implant dentistry have reported accelerated soft-tissue healing as well as osseointegration. In addition, it was suggested that the ultrasound treated wounds were at a more advanced stage in the repair process¹². The cellular mechanisms underlying LIPUS induced tissue regeneration was done and studies by Ikai et al. (2008) showed that a daily LIPUS treatment protocol of 20 minutes for a period of 4 weeks has a beneficial effect on gingival epithelium cells, accelerating periodontal wound healing after flap surgery⁸. In other study using gingival epithelial cells, Shiraishi et al (2011) reported that LIPUS accelerates soft-tissue healing by increasing the expression of connective tissue growth factor (CCN2/CTGF), an important gene for wound healing and angiogenesis in periodontal tissues⁹.

Lipus and Implant Osseointegration

The use of endosseous dental implants for replacing missing teeth increased considerable over the last few years and may be considered as the current most popular treatment option for edentulous patients. LIPUS helps in enhancement of endogenous bone healing around biomaterials through different forms of biophysical stimulations²³. A study done by Tanzer et al. (1996) showed that LIPUS enhanced the rate and extent of bone growth into fully porous coated implants inserted into dog femora²⁴. Studies done by Hsu et al. (2011) showed that ultrasound stimulation helps in good blood flow and

mature type I collagen fibers around titanium implants, and accelerated bone formation. In addition, pulsed ultrasound effectively promotes cell migration and new bone regeneration in tissue culture of MG63 osteoblast like cells²⁵. Studies done by Ustun et al. (2008) showed that LIPUS stimulation increases the area, bone volume and bone implant contact ratio values in tibial bone suggesting that LIPUS application may accelerate and promote bone healing around dental implants leading to a higher quality and faster osseointegration²⁶.

Lipus and Gene Delivery

Periodontal disease or inflammatory root resorption is relevant pathologic condition that can lead to tooth loss. In this regard, several tissue-engineering techniques have been proposed to restore periodontal integrity²⁷. Administration of growth factors has proved to exert positive effect with some clinical limitations such as proteolytic degradation, rapid diffusion, and solubility of the delivery vehicle²⁸. In order to optimize the results, gene transfer methods were introduced with limitation of progress in clinical periodontal gene therapy by the immunogenicity and cytotoxicity of viral vectors and low transfection efficiency with regard to non-viral vectors²⁹. Therapeutic ultrasound associated to echo contrast agents such as nano/microbubbles can optimize gene transfection in vitro and in vivo³⁰. Recently, it has been shown that therapeutic ultrasound also provides an effective gene delivery system for bone and periodontal regeneration³¹. Studies done by Watanuki et al. (2009) showed that LIPUS stimulation in mouse calf muscles injected with BMP-4 plasmids and transcutaneous electroporated showed increased ectopic calcium and total collagen content and bone area³².

Discussion

Among the causes of teeth loss, inflammatory root resorption has received a great concern due to its unpredictability, difficult control and lack of biological understanding. The resorption of hard tissue in primary teeth is a normal physiologic phenomenon. On the other hand, the hard tissues of permanent teeth are not resorbed under healthy conditions and the resorption is thus considered a pathologic process³³. Different causes have been attributed to the root resorption process including pressure, inflammation, neoplastic process and systemic conditions³⁴. The major problem encountered after tooth replantation and orthodontic tooth movement is the resorption of root. Approaches aiming to inhibit root resorption and restore periodontal integrity during the dental practice are still the subject of debate and investigation. Some therapeutic approaches have been proposed to inhibit root resorption after tooth replantation and can induce periodontal regeneration³⁵. In this context, non-invasive modality such as LIPUS therapy has been given increased attention and risen as promising therapeutic tool for the regeneration of periodontium. The effectiveness of LIPUS for bone regeneration is already universally accepted and some recent papers have provided evidence that it can exert beneficial effects in other kinds of tissues, including teeth. In addition, LIPUS

presents low toxicity, low immunogenicity, non-invasiveness, highly targeted selectivity, and repeated applicability. However, the diversity of techniques, application protocols and ultrasound specifications found in the literature may cause confusion for the clinician. Bains et al. (2008) have pointed out that ultrasound application in both diagnosis and periodontal therapy seems to present promising results; however, long-term evidence-based studies are required to use ultrasound in routine periodontal practice³⁶.

Conclusion

Despite LIPUS therapy has been widely used in the fields of orthopedic surgery and rehabilitation, its availability by dental professionals is still incipient. The effects of LIPUS in bony tissue seem to be well understood, but the literature has still lacked for available information about its effects on periodontal tissues. The present review brings out current evidence that LIPUS has a positive effect on tooth and periodontal cells metabolism, suggesting that LIPUS can be a promising therapeutic tool for the regeneration of tooth support tissues.

References

- Petrtyl M, Hert J, Fiala P. Spatial organization of the haversian bone in man. *J Biomech* 1996; 29: 1619.
- Tan SD, Xie R, Klein-Nulend J, Van Rheden RE, Bronckers AL, Kuijpers-Jagtman AM, Van den Hoff JW, Maltha JC. Orthodontic force stimulates eNOS and iNOS in rat osteocytes. *J Dent Res* 2009; 88: 255-60.
- Buckley MJ, Banes AJ, Levin LG, et al. Osteoblasts increase their rate of division and align in response to cyclic mechanical tension in vitro. *Bone Miner* 1998; 4: 225-36.
- Claes L, Willie B. The enhancement of bone regeneration by ultrasound. *Prog Biophys Mol Biol* 2007; 93: 384-98.
- Khan Y, Laurencin CT. Fracture repair with ultrasound: clinical and cell-based evaluation. *J Bone Joint Surg* 2008; 90: 138-44.
- Duarte LR. The stimulation of bone growth by ultrasound. *Arch Orthop Trauma Surg* 1983; 101: 153-9. [14] Dyson M. Therapeutic applications of ultrasound. In: Biological effects of ultrasound. Nyborg WL, Ziskin MC, Eds. Churchill Livingstone, New York, NY 1985; pp. 121-33.
- Chen YJ, Wang CJ, Yang KD, et al. Pertussis toxin-sensitive Gα protein and ERK dependent pathways mediate ultrasound promotion of osteogenic transcription in human osteoblasts. *FEBS Lett* 2003; 554: 154-8.
- Naruse K, Miyauchi A, Itoman M, Mikuni-Takagaki Y. Distinct anabolic response of osteoblasts to low-intensity pulsed ultrasound. *J Bone Miner Res* 2003; 18: 360-9.
- Kokubu T, Matsui N, Fujioka H, Tsunoda M, Mizuno K. Low intensity pulsed ultrasound exposure increases prostaglandin E2 production via the induction of cyclooxygenase-2 mRNA in mouse osteoblasts. *Biochem Biophys Res Commun* 1999; 256: 284-7.
- Bosshardt DD, Degen T, Lsng NP. Sequence of protein expression of bone sialoprotein and osteopontin at the developing interface between repair cementum and dentin in human deciduous teeth. *Cell Tissue Res* 2005; 320: 399-407.
- Matsuda N, Yokoyama K, Takeshita, S, Watanabe M. Role of epidermal growth factor and its receptor in mechanical stress-induced differentiation of human periodontal ligament cells in vitro. *Arch Oral Biol* 1998; 43: 987-97.
- Young SR, Dyson M. The effect of therapeutic ultrasound on angiogenesis. *Ultrasound Med Biol* 1990; 16: 261-9. 224 The Open Dentistry Journal, 2012, Volume 6 Braga Rego et al.
- Inubushi T, Tanaka E, Rego EB, et al. Effects of ultrasound on the proliferation and differentiation of cementoblast lineage cells. *J Periodontol* 2008; 79: 984-90.
- Mostafa NZ, Uludağ H, Dederich DN, Doschak MR, El-Bialy TH. Anabolic effects of low-intensity pulsed ultrasound on human gingival fibroblasts. *Arch Oral Biol* 2009; 54: 743-8.
- Ten Cate AR. The periodontium: Oral histology, development, structure and function. Mosby, St Louis, MO 2003; 276-9.
- Hollender L, Ronneman A, Thilander B. Root resorption, marginal bone support and clinical crown length in orthodontically treated patients. *Eur J Orthod* 1980; 2: 197-205.
- Pavlin D, Gluhak J. Effect of mechanical loading on periodontal cells. *Crit Rev Oral Biol Med* 2001; 12: 414-24.
- El-Bialy T, El-Shamy I, Graber TM. Repair of orthodontically induced root resorption by ultrasound in humans. *Am J Orthod Dentofacial Orthop* 2004; 126: 186-93.
- Dalla-Bona DA, Tanaka E, Oka H, et al. Effects of ultrasound on cementoblast metabolism in vitro. *Ultrasound Med Biol* 2006; 32: 943-8.
- Rego EB, Inubushi T, Kawazoe A, et al. Ultrasound stimulation induces PGE2 synthesis promoting cementoblastic differentiation through EP2/EP4 receptor pathway. *Ultrasound Med Biol* 2010; 36: 907-15.
- Rego EB, Inubushi T, Miyauchi M, et al. Ultrasound stimulation attenuates root resorption on rat replanted molars and impairs TNF-α signaling in vitro. *J Periodont Res* 2011; 46: 648-54.
- Bränemark PI, Zarb G, Albrektsson T. Tissue-integrated prosthesis. In: Bränemark PI, editor. Osseointegration in clinical dentistry. Chicago, IL: Quintessence Publishing Co; 1985; pp. 11-76..
- Fini M, Giavarelli G, Setti S, Martini L, Torricelli P, Giardino R. Current trends in the enhancement of biomaterial osteointegration: Biophysical stimulation. *Biomaterials* 2004; 27: 681-90.
- Tanzer M, Harvey E, Kay A, Morton P, Bobyn JD. Effect of non-invasive low intensity ultrasound on bone growth into porous-coated implants. *J Orthop Res* 1996; 14: 901-6.
- Hsu SK, Huang WT, Liu BS, Li SM, Chen HT, Chang CJ. Effects of near-field ultrasound stimulation on new bone formation and osseointegration of dental titanium implants in vitro and in vivo. *Ultrasound Med Biol* 2011; 37: 403-16.
- Ustun Y, Erdogan O, Kurkcu M, Akova T, Damlar I. Effects of low-intensity pulsed ultrasound on dental implant osseointegration: a preliminary report. *Eur J Dent* 2008; 2: 254-62.
- Ramseier CA, Abramson ZR, Jin Q, Giannobile WV. Gene therapeutics for periodontal regenerative medicine. *Dent Clin North Am* 2006; 50: 245-63.
- Pradeep AR, Karthikeyan BV. Tissue engineering: prospect for regenerating periodontal tissues. *Indian J Dent Res* 2003; 14: 224-9.
- Ramseier CA, Abramson ZR, Jin Q, Giannobile WV. Gene therapeutics for periodontal regenerative medicine. *Dent Clin North Am* 2006; 50: 245-63.
- Nishida K, Doita M, Takada T, et al. Sustained transgene expression in intervertebral disc cells in vivo mediated by microbubble-enhanced ultrasound gene therapy. *Spine* 2006; 31: 1415-9.
- Sheyn D, Kimelman-Bleich N, Pelleg G, Zilberman Y, Gazit D, Gazit Z. Ultrasound-based nonviral gene delivery induces bone formation in vivo. *Gene Ther* 2008; 15: 257-66.
- Watanuki M, Kishimoto KN, Kotajima S, Iwabuchi S, Kokubun S. Effect of low-intensity pulsed ultrasound on scaffold-free ectopic bone formation in skeletal muscle. *Ups J Med Sci* 2009; 114: 242-8.
- Gunraj MN. Dental root resorption. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1999; 88: 647-53.
- Ravindran S, Chaudhary M, Tumsare M, Patil S, Wadhwan V. A scanning electron microscopic study of the patterns of external root resorption under different conditions. *J Appl Oral Sci* 2009; 17: 481-6.
- Loberg EL, Engstrom H. Thyroid administration to reduce root resorption. *Angle Orthod* 1994; 64: 395-9.
- Bains VK, Mohan R, Bains R. Application of ultrasound in periodontics: Part II. *J Indian Soc Periodontol* 2008; 12: 55-61.